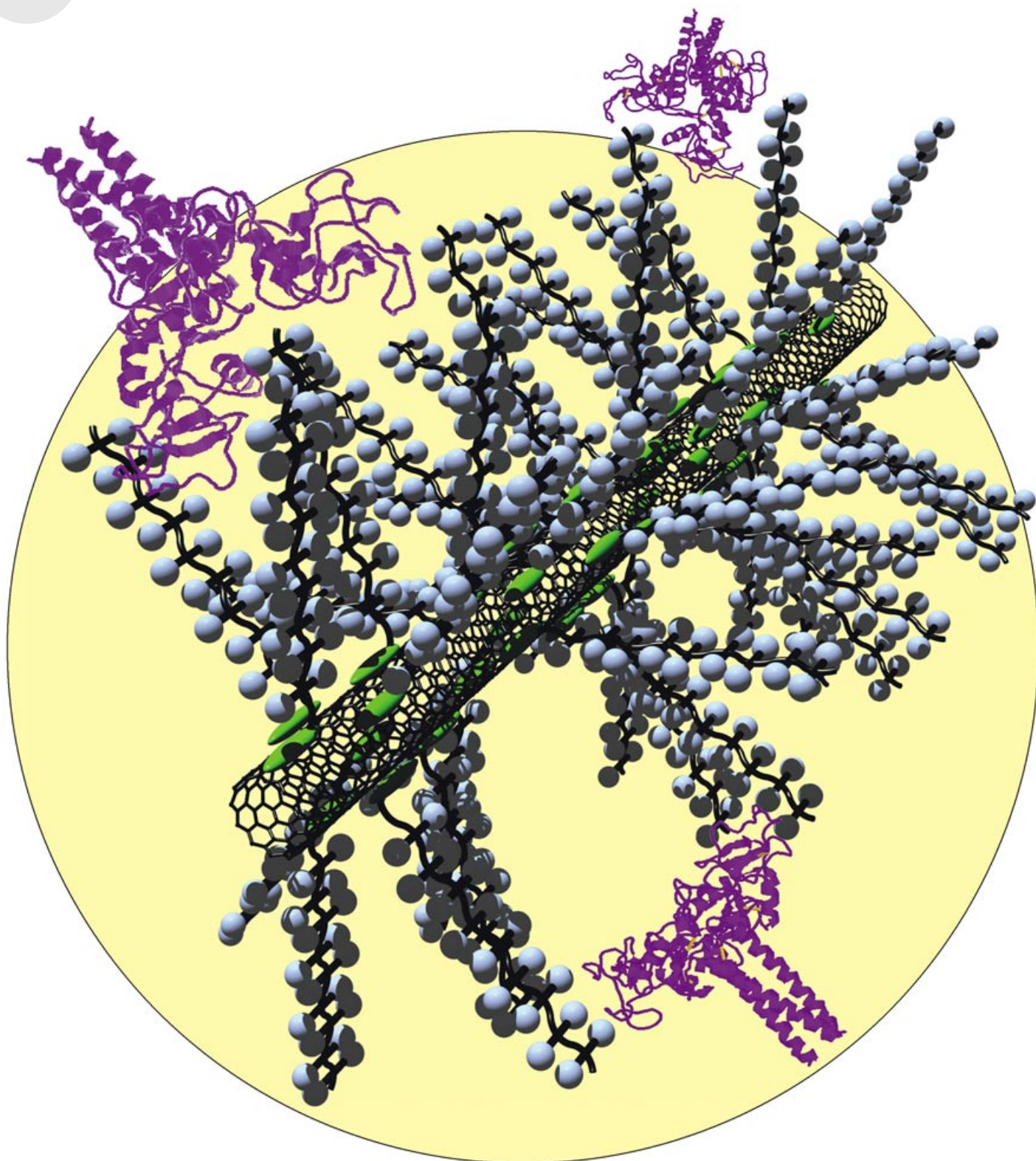


Communications



Carbohydrate-functionalized polymers designed to mimic the structures of mucin glycoproteins assembled on carbon nanotubes are water soluble and can be functionalized with ligands for specific receptor binding. For more information see the Communication by A. Zettl, C. R. Bertozzi et al. on the following pages.

Biomimetic Synthesis

Biomimetic Engineering of Carbon Nanotubes by Using Cell Surface Mucin Mimics**

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One of the most exciting applications of nanoscale science and technology is in the exploration of biological systems.^[1] Carbon nanotubes (CNTs) have unique structural, mechanical, and electrical properties, and consequently numerous potential applications in biology, including sensing,^[2] imaging,^[3] and scaffolding for cell growth.^[4] Few of these applications have yet been realized, however, because of the incompatibility of the CNT surface, which is hydrophobic and prone to nonspecific bioadsorption, with biological components such as cells and proteins. In addition, the aqueous environment required for biological materials are not suitable for unfunctionalized CNTs.^[5] In nature, cells are faced with a similar challenge of resisting nonspecific biomolecule interactions while engaging in specific molecular recognition. These functions can be simultaneously fulfilled by mucin glycoproteins, defined by their dense clusters of O-linked glycans. Here we describe a biomimetic surface modification of CNTs using glycosylated polymers designed to mimic natural cell-surface mucins. CNTs modified with mucin mimics were soluble in water, resisted nonspecific protein binding, and bound specifically to biomolecules through receptor–ligand interactions. This strategy for biomimetic surface engineering provides a means to bridge nanomaterials and biological systems.

Mucins coat the surfaces of numerous cell types^[6] and present epitopes for receptor-mediated cell–cell recogni-

tion.^[7] Their dense glycosylation confers rigidity to the polypeptide backbone and thereby extends the mucin polymer well above the cell surface.^[8] In addition, the glycans provide for strong hydration and passivation against biofouling. We have recently developed glycosylated polymers that share many properties with natural mucins (Figure 1 a).^[9] In native mucins, the clustered α -N-acetylgalactosamine (α -GalNAc) residues proximal to the peptide are the major contributors to the rigidification of peptide backbones. Although the core α -GalNAc residue is usually elaborated with additional sugars, removal of those sugars does not affect the mucin architecture.^[10] The importance of the α -GalNAc residue for mucin structure has also been suggested by NMR analysis of synthetic glycopeptides.^[11] Accordingly, we designed a synthetically tractable mucin mimic in which α -GalNAc residues were linked through an oxime bond to a poly(methyl vinyl ketone) [poly(MVK)] backbone (Figure 1 a). The synthesis involved chemoselective ligation of poly(MVK) with an aminooxy-functionalized GalNAc analogue.^[12] Light-scattering analysis indicated that the α -GalNAc-conjugated polymers adopt a rigid extended structure in water, similar to native mucin, whereas the unconjugated polymers adopt a more conventional globular structure.^[9]

We introduced a C_{18} lipid at one end of a mucin mimic polymer with a molecular weight of about 75000 g mol^{-1} to enable surface modification of CNTs (Figure 1 b; see Supporting Information for synthetic details). The hydrated diameter of the lipid-terminated mucin mimic was determined by light-scattering analysis to be about 54 nm. Lipids are known to self-assemble on the surface of CNTs through hydrophobic interactions in the presence of water^[13] and lipid-functionalized glycopolymers have been shown to form ordered arrays on graphite surfaces.^[14] We envisioned that lipid-functionalized mucin mimics could assemble on CNTs in a similar manner as the organization of native mucins in the cell membrane, with the glycosylated polymers projecting into the aqueous medium (Figure 1 c).

We first subjected single-walled carbon nanotubes (SWNTs) to ultrasonication in the presence of an aqueous solution of the C_{18} -functionalized mucin mimic bearing α -GalNAc residues (C_{18} - α -MM, Figure 1 b). The SWNTs were fully solubilized during the procedure (Figure 2 a), which suggests the formation of a hydrophilic surface coating. A similar procedure was applied to multiwalled carbon nanotubes (MWNTs) with the same outcome (Figure 2 c). The suspensions of C_{18} - α -MM-functionalized CNTs were stable for more than three months, while unfunctionalized CNTs precipitated very quickly (within hours) in aqueous solutions (Figure 2 b and d). The C_{18} lipid on the mucin mimic polymer was essential to solubilize the CNTs: they rapidly precipitated from solutions of polymers identical to the above mucin mimics but lacking the lipid tail (not shown). These observations are consistent with a model in which C_{18} - α -MMs coat the CNTs as shown in Figure 1 c.

The CNTs coated with mucin mimics (C_{18} - α -MM-SWNTs, for example) were directly characterized by atomic force microscopy (AFM), scanning electron microscopy (SEM), and transmission electron microscopy (TEM). As produced,

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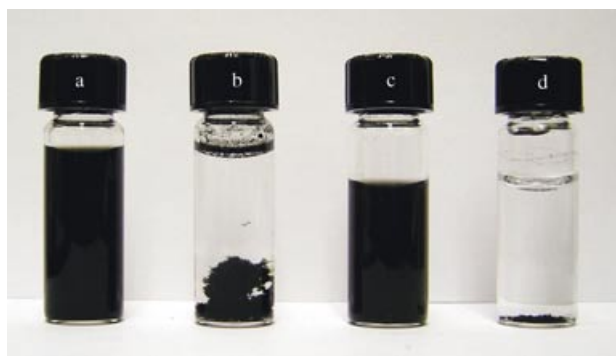


Figure 2. Photographs of vials containing a) a stable suspension of $C_{18}\text{-}\alpha\text{-MM-SWNTs}$ in H_2O after more than three months, b) as-produced SWNTs in H_2O , which precipitate in several hours, c) stable suspension of $C_{18}\text{-}\alpha\text{-MM-MWNTs}$ in H_2O after more than three months, d) as-produced MWNTs in H_2O , which precipitate over several hours.

individual nanotubes, as observed by all three imaging techniques. AFM permitted analysis of the modified tubes at ambient pressure and in an hydrated form, which is important for maintaining the structure of the mucin coating. As shown in Figure 3a, tapping mode AFM images on a silicon substrate revealed $C_{18}\text{-}\alpha\text{-MM-SWNTs}$ with fairly uniform diameters of 65–70 nm. The idealized model shown in Figure 1c would predict a diameter on the order of 100 nm,

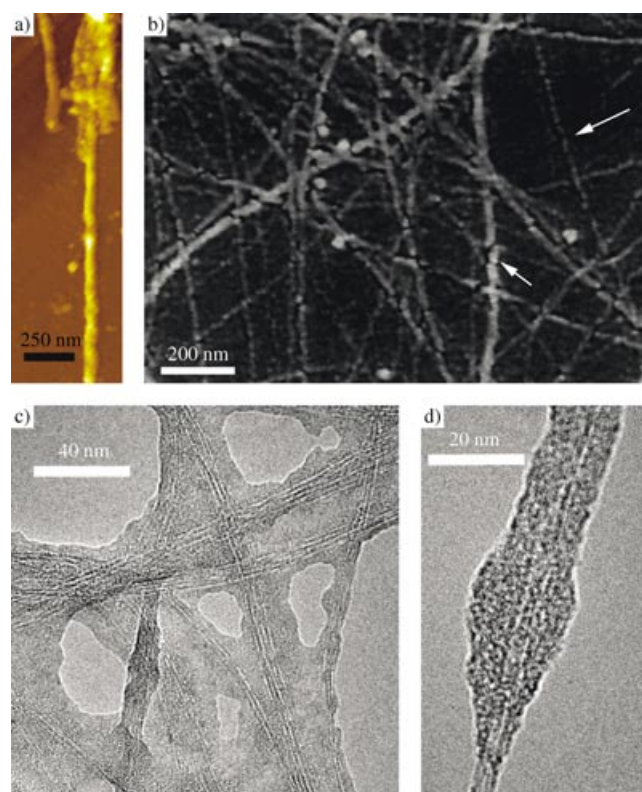


Figure 3. a) Tapping mode AFM image of $C_{18}\text{-}\alpha\text{-MM-SWNTs}$ on a silicon substrate. b) SEM image of $C_{18}\text{-}\alpha\text{-MM-SWNTs}$. The arrows indicate damaged regions of the mucin mimic coating caused by the electron beam. c), d) TEM images of $C_{18}\text{-}\alpha\text{-MM-SWNTs}$ stained with 0.5 % methylamine vanadate.

roughly twice that of the polymer mimicking the mucin (the SWNT itself is only ca. 2 nm in diameter). However, distortions to the soft mucin coating imposed by the AFM tip or substrate, or a nonperpendicular angle of projection of the mucin polymer from the SWNT surface could account for the discrepancy.

SEM analysis of $C_{18}\text{-}\alpha\text{-MM-SWNTs}$ also provided evidence of the mucin coating. Exposure of the sample to the 5-keV electron beam led to visible sites of damage along the SWNT surface, as indicated by the arrows in Figure 3b. This result was expected based on the sensitivity of organic species to decomposition induced by an electron beam. The damage induced by the electron beam increased with longer exposure times, and culminated in near-complete destruction of the coating after 15 minutes (see the Supporting Information). By contrast, unmodified CNTs are fairly stable under the electron beam, and showed no visible change in surface morphology after prolonged exposure.

TEM analysis of $C_{18}\text{-}\alpha\text{-MM-SWNTs}$ provided a direct visualization of the mucin coating (Figure 3c and d). The thickness of the coating varied from 10 to 25 nm under these conditions, in which the sample is dehydrated under high vacuum. The coating was associated with the CNT surface (Figure 3c), and was not observed in regions of the image lacking CNTs. A single tube (or small bundle) coated with $C_{18}\text{-}\alpha\text{-MM}$ is shown in Figure 3d. The coating was not entirely uniform along the length of this SWNT, perhaps the result of damage induced by the electron beam (100 keV) or collapse of the mucin polymers under high vacuum. In this regard, AFM imaging is a superior technique for visualizing mucin-coated SWNTs as they would exist in a functionally relevant aqueous environment.

On cells, mucins serve the dual role of molecular recognition and resistance to biofouling. We sought to determine if these same functions could be realized in the context of the nanotube surface. The $C_{18}\text{-}\alpha\text{-MM}$ coating introduces $\alpha\text{-GalNAc}$ residues onto the CNT surface, which could be recognized by an $\alpha\text{-GalNAc}$ -specific receptor such as the lectin *Helix pomatia* agglutinin (HPA).^[15] To test this we incubated $C_{18}\text{-}\alpha\text{-MM-SWNTs}$ with a solution of HPA conjugated with fluorescein isothiocyanate (HPA-FITC; shown schematically in Figure 4a I). The $C_{18}\text{-}\alpha\text{-MM-SWNTs}$ were dialyzed to remove excess HPA-FITC and then analyzed for bound lectin by fluorescence spectroscopy. The $C_{18}\text{-}\alpha\text{-MM-SWNTs}$ showed significant fluorescence, which was attributed to bound fluorescein (trace I of Figure 4b). HPA-FITC labeling of the $C_{18}\text{-}\alpha\text{-MM-SWNTs}$ was inhibited when 0.2 M free GalNAc was present in solution (Figure 4a III and trace III of Figure 4b), thus confirming that fluorescent labeling was dependent on the receptor–ligand interaction.

In addition, we prepared a similar mucin mimic in which the GalNAc residues were conjugated to the polymer backbone through a β -anomeric linkage ($C_{18}\text{-}\beta\text{-MM}$; see Supporting Information for synthetic details). This mucin mimic was physically identical to its α -linked counterpart but should not be capable of binding HPA.^[16] We coated SWNTs with $C_{18}\text{-}\beta\text{-MM}$ and incubated them with HPA-FITC (shown schematically in Figure 4a II). No significant fluorescence labeling was observed (trace II in Figure 4b), thus indicating that the lectin

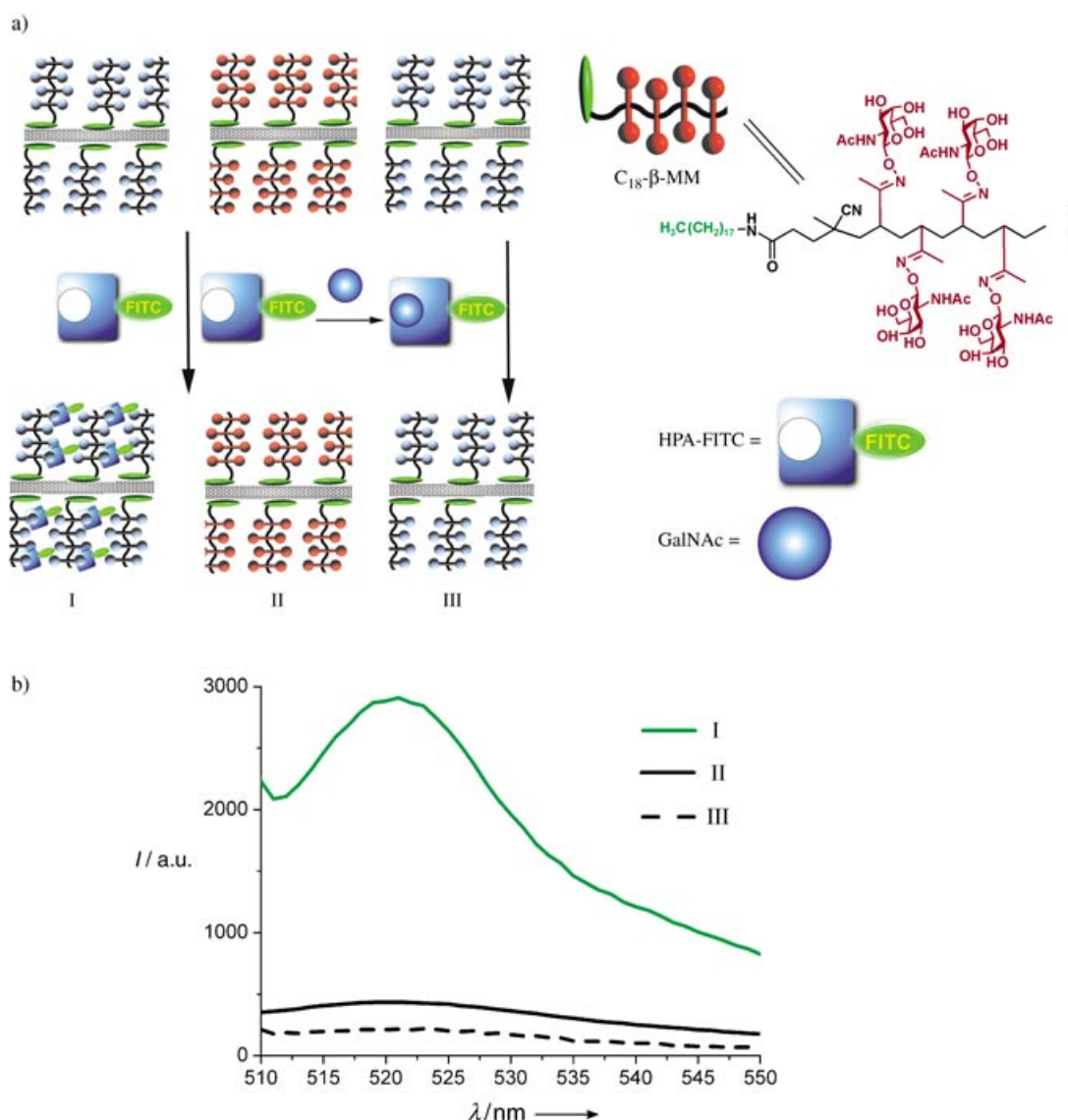


Figure 4. Specific binding of HPA to C₁₈-α-MM-coated nanotubes. a) Scheme for: I) specific binding of HPA to the surface of C₁₈-α-MM-SWNTs, II) lack of binding of HPA to C₁₈-β-MM-SWNTs, III) inhibition of HPA binding by soluble GalNAc. b) Fluorescence spectra (510–550 nm) showing the specific binding of HPA to the surface of C₁₈-α-MM-SWNTs. Trace I: spectrum showing bound HPA-FITC on the surface of C₁₈-α-MM-SWNTs; trace II: spectrum showing the lack of bound HPA to C₁₈-β-MM-SWNTs; trace III: spectrum showing fluorescence associated with HPA-FITC binding in the presence of soluble GalNAc. The excitation wavelength was 492 nm. Spectra were corrected for background fluorescence by subtracting the fluorescence spectrum of C₁₈-α-MM-SWNTs or C₁₈-β-MM-SWNTs alone. These data are representative of results observed in triplicate experiments.

does not interact with the tubes coated with the mucin mimic in the absence of its preferred ligand. These results demonstrate that CNTs coated with a mucin mimic can engage in specific molecular recognition with protein receptors and resist nonspecific protein binding.

In summary, we have developed a practical and general approach to engineering biomimetic surfaces on CNTs. The mucin mimics used in this study endowed the CNT surfaces with properties shared by cell surfaces, including the display of carbohydrates capable of molecular recognition. The synthetic process leading to the mucin mimic allows for the introduction of myriad functional epitopes, in addition to sugars, that could encode interactions with numerous receptor

types. This work should facilitate the integration of CNTs into aqueous biological systems that include proteins and cells.

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- [1] a) M. Sarikaya, C. Tamerler, A. K. Y. Jen, K. Schulten, F. Baneyx, *Nat. Mater.* **2003**, 2, 577; b) G. M. Whitesides, *Nat. Biotechnol.* **2003**, 21, 1161; c) P. Alivisatos, *Nat. Biotechnol.* **2004**,

- 22, 47; d) D. H. Reich, M. Tanase, A. Hultgren, L. A. Bauer, C. S. Chen, G. J. Meyer, *J. Appl. Phys.* **2003**, 93, 7275.
 - [2] R. J. Chen, S. Bangsaruntip, L. A. Drouvalakis, N. Wong, S. Kam, M. Shim, Y. Li, W. Kim, P. J. Utz, H. Dai, *Proc. Natl. Acad. Sci. USA* **2003**, 100, 4984.
 - [3] S. S. Wong, E. Joselevich, A. T. Woolley, C. L. Cheung, C. M. Lieber, *Nature* **1998**, 394, 52.
 - [4] M. P. Mattson, R. C. Haddon, A. M. Rao, *J. Mol. Neurosci.* **2000**, 14, 175.
 - [5] Y. Lin, S. Taylor, H. P. Li, K. A. S. Fernando, L. W. Qu, W. Wang, L. R. Gu, B. Zhou, Y. P. Sun, *J. Mater. Chem.* **2004**, 14, 527.
 - [6] G. J. Strous, J. Dekker, *Crit. Rev. Biochem. Mol. Biol.* **1992**, 27, 57.
 - [7] Y. Shimizu, S. Shaw, *Nature* **1993**, 366, 630.
 - [8] J. Hikens, M. J. L. Ligtenberg, H. L. Vos, S. V. Litvinov, *Trends Biochem. Sci.* **1992**, 17, 359.
 - [9] G. S. Lee, Y. Shin, I. Choi, H. Hahn, C. R. Bertozzi, *J. Am. Chem. Soc.*, submitted.
 - [10] a) R. Shogren, T. A. Gerken, N. Jentoft, *Biochemistry* **1989**, 28, 5525; b) T. A. Gerken, K. J. Butenhof, R. Shogren, *Biochemistry* **1989**, 28, 5536.
 - [11] D. H. Live, L. J. Williams, S. D. Kuduk, J. B. Schwarz, P. W. Glunz, X. T. Chen, D. Sames, R. A. Kumar, S. J. Danishefsky, *Proc. Natl. Acad. Sci. USA* **1999**, 96, 3489.
 - [12] L. A. Marcaurelle, Y. Shin, S. Goon, C. R. Bertozzi, *Org. Lett.* **2001**, 3, 3691.
 - [13] C. Richard, F. Balavoine, P. Schultz, T. W. Ebbesen, C. Mioskowski, *Science* **2003**, 300, 775.
 - [14] N. B. Holland, Y. Qiu, M. Ruegsegger, R. E. Marchant, *Nature* **1998**, 392, 799.
 - [15] N. Sharon, *Adv. Immunol.* **1983**, 34, 213.
 - [16] V. Piller, F. Piller, J. Cartron, *Eur. J. Biochem.* **1990**, 191, 461.
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